



**SoilWealth | ICP**  
nurturing crops · protecting crops

The role of soil DNA and traditional pathology.

– how is it being used and what can it do for you?

Michael Rettke  
SARDI



# Soilborne disease diagnostics

**Traditional**

**Molecular**



Traditional	Molecular
<b>Strengths</b>	<b>Strengths</b>
Can identify wide range pathogens / disorders	Highly specific
Does not always require specialist equipment	Quantification (including beneficials)
Can incorporate other factors	Test for many pathogens at once
<b>Limitations</b>	<b>Limitations</b>
Requires knowledge, experience and skill	Requires specialist equipment
Time consuming and soilborne diseases can be challenging	New pathogen tests require validation (long process)
Accuracy	Relies on accuracy and availability of DNA sequence for pathogens





# Investigative



Post mortem to inform future decisions

# Predictive (risk)



Inform decisions to avoid a post mortem



# Investigative diagnostics – Uses

- **New problem**
  - ❑ **Identification of cause**
- Problem that increasing
  - ❑ Identification of cause
  - ❑ Assessment of resistance to controls
- Industry unsure or questioning the cause
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# Investigative diagnostics the process



**Visual examination**



**Microscopic observation / Test kits**



**Isolation of pathogen**



**Identification**



**Pathogenicity test**

**(Nematodes are normally quantified)**



# What pathogens are in this root system?



Pathogen	Disease
<i>Plasmodiophora brassicae</i>	Club root
<i>Rhizoctonia solani</i> (AG 2.1, 4)	Rhizoctonia
<i>Plasmodiophora brassicae</i>	Club root
<i>Verticillium dahliae</i>	Verticillium wilt
<i>Leptosphaeria maculans</i>	Blackleg
<i>Pythium</i> (Clade F, Clade I)	Pythium
<i>Pratylenchus</i> spp.	Root lesion nematodes
<i>Meloidogyne</i> spp.	Root knot nematodes

# Report on pathogen levels



Pathogen	Disease		Detection
<i>Plasmodiophora brassicae</i>	Club root	High	9780126 <sub>kDNA</sub>
<i>Rhizoctonia solani</i> (AG 2.1)	Rhizoctonia	High	27711 <sub>pgDNA</sub>
<i>Rhizoctonia solani</i> (AG 4)	Rhizoctonia		Below detection
<i>Verticillium dahliae</i>	Verticillium wilt		Below detection
<i>Leptosphaeria maculans</i>	Blackleg	High	20408 <sub>pgDNA</sub>
<i>Pythium</i> (Clade F)	Pythium	Low	54 <sub>pgDNA</sub>
<i>Pythium</i> (Clade I)	Pythium	Mod	1043 <sub>pgDNA</sub>
<i>Pratylenchus</i> spp.	Root lesion nematodes		Below detection
<i>Meloidogyne javanica/incognita/arenaria</i>	Root knot nematodes	Low	14 <sub>pgDNA</sub>



# Investigative diagnostics – Utilisation

- Most state agencies and several specialist laboratories offer diagnostic services
- Most samples from commercial horticulture operations are the difficult ones or disease complexes
- Critical part of research and development projects

**If do not know the cause  
and contributors to risk  
– how can it be managed**

# Disease risk assessment – Uses

## Testing services

- Identify disease risk prior to planting
- In crop monitoring
- Assess storage potential
- Monitor impact changed practices



# Identify disease risk prior to planting

## Traditional

- Seed testing
- Soil baiting
- Water testing
- Nematode counts
- Sieving / Floatation / Extraction and quantification
- Bioassay

**Low availability of disease risk assessment based on traditional techniques**

## Exception is pre-plant nematode testing

- Established techniques for nematode counting
- Established relationships between pre-plant nematode population and risk of crop damage





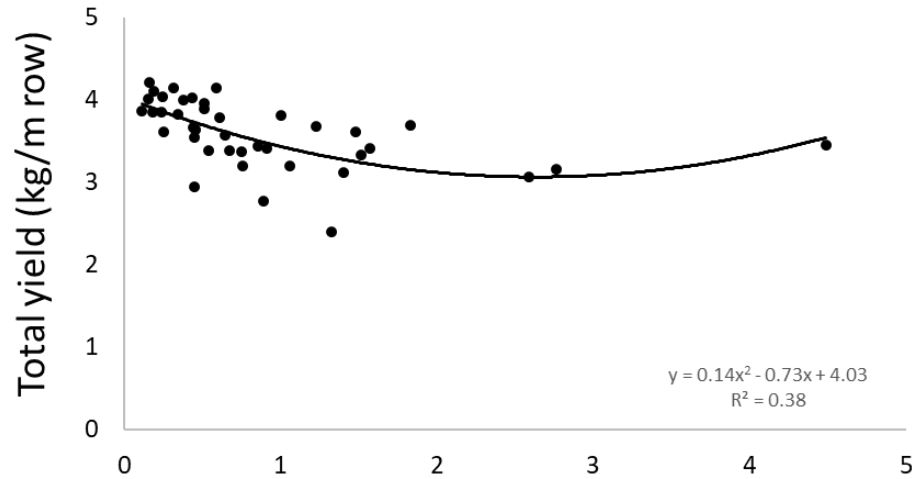
# Identify disease risk prior to planting

## Molecular

- Relatively fast
- High throughput
- Assess multiple targets
- Quantitative



# Identify disease risk prior to planting



Soil inoculum DNA *Pratylenchus thornei* (nematodes/g)

## Yield loss in carrots

Relatively low populations of root lesion nematodes associated with loss of yield



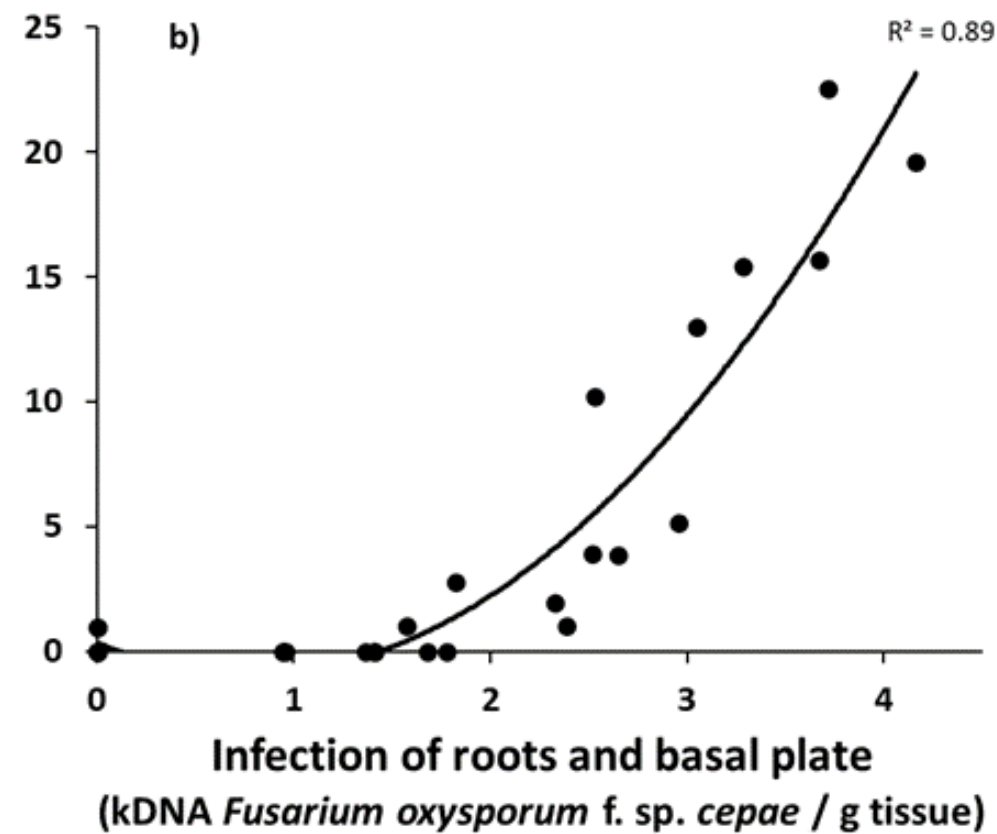
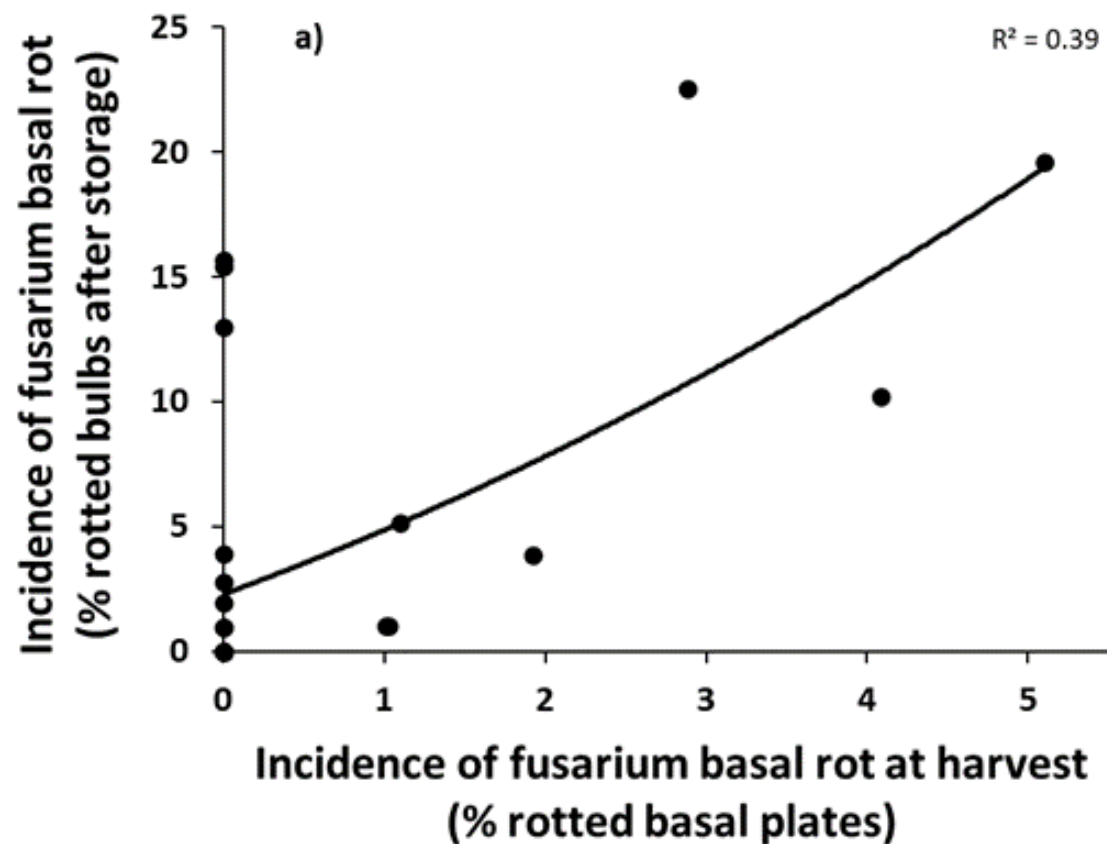
# Assess storage potential



Visual observation



DNA testing





# What can we detect – Hort Veg DNA tests

## NEMATODES

**Root knot**  
(5 species)

**Root lesion**  
(5 species)

**Stem**  
(1 species)

**Free living**  
(15 tests)

## FUNGI

***Rhizoctonia***  
(5 tests)

***Pythium***  
(4 tests)

**Other**  
(10 species)

**Beneficial**  
(8 tests)

## BACTERIA

***Streptomyces***  
(1 test)

## OTHER

***Plasmodiophora***  
(1 test)

Tests relevant to  
vegetable industry

[http://www.pir.sa.gov.au/research/services/molecular\\_diagnostics](http://www.pir.sa.gov.au/research/services/molecular_diagnostics)



# Commercial service – Potato



Sample: **APA**

Grower:   
 Paddock:   
 Sample point:   
 Nearest town:

**Molecular Diagnostic Centre**  
 Gate 2A, Hartley Grove  
 URRBRAE SA 5064  
 P 08 8429 2284

Report date:   
 Date sampled:   
 Dry weight (g): **436**  
 Sample condition: **Damp**  
 Sample no.: **1 of 1**  
 Sample taken: **Pre-plant Not cultivated**  
 Order No.:

Paddock history **Years since last potato crop 5 Years**

TEST	RESULT	DISEASE RISK*			
		Not Detected	Low	Med	High
Colletotrichum coccodes	2.0 log(pg DNA/g soil)	Not Detected	Low	Med	High
Verticillium dahliae	<0.1 log(pg DNA/g soil)	Not Detected	Low	Med	High
Spongospora subterranea	1.5 log(pg DNA/g soil)	Not Detected	Low	Med	High
Meloidogyne fallax	2.3 log(pg DNA/g soil)	Not Detected	Low	Med	High
Meloidogyne javanica/incognita/arenaria	<0.1 log(pg DNA/g soil)	Not Detected	Low	Med	High

\*Risk categories should be used as a guide only, may be subject to regional and seasonal differences, and may be revised over time.

TEST	RESULT	POPULATION DENSITY**			
		Not Detected	Low	Med	High
Helminthosporium solani	<0.1 log(kDNA copies/g soil)	Not Detected	Low	Med	High
Rhizoctonia solani AG2.1	1.5 log(pg DNA/g soil)	Not Detected	Low	Med	High
Rhizoctonia solani AG3	<0.1 log(pg DNA/g soil)	Not Detected	Low	Med	High
Rhizoctonia solani AG4	<0.1 log(pg DNA/g soil)	Not Detected	Low	Med	High
Sclerotinia sclerotiorum/S. minor	0.5 log(kDNA copies/g soil)	Not Detected	Low	Med	High
Phytophthora erythroseptica	<0.1 log(kDNA copies/g soil)	Not Detected	Low	Med	High
Pythium clade I	0.4 log(pg DNA/g soil)	Not Detected	Low	Med	High
Streptomyces txtA gene	<0.1 log(pg DNA/g soil)	Not Detected	Low	Med	High
Pratylenchus crenatus	<0.1 nematodes/g soil	Not Detected	Low	Med	High
Pratylenchus neglectus	1.2 nematodes/g soil	Not Detected	Low	Med	High
Pratylenchus penetrans	<0.1 nematodes/g soil	Not Detected	Low	Med	High
Meloidogyne hapla	<0.1 log(pg DNA/g soil)	Not Detected	Low	Med	High

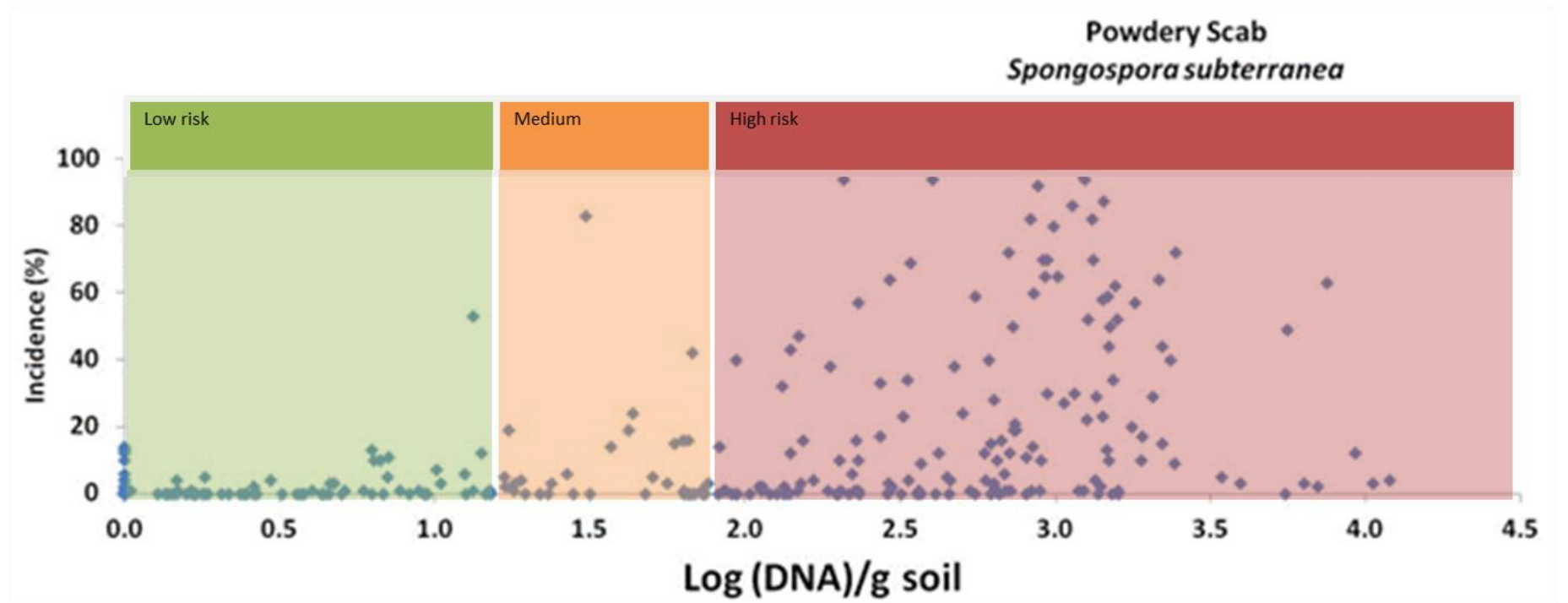
\*\*Population densities are based on the pathogen levels detected in PREDICTA samples across the potato industry. These are not disease risk categories.



# Commercial service



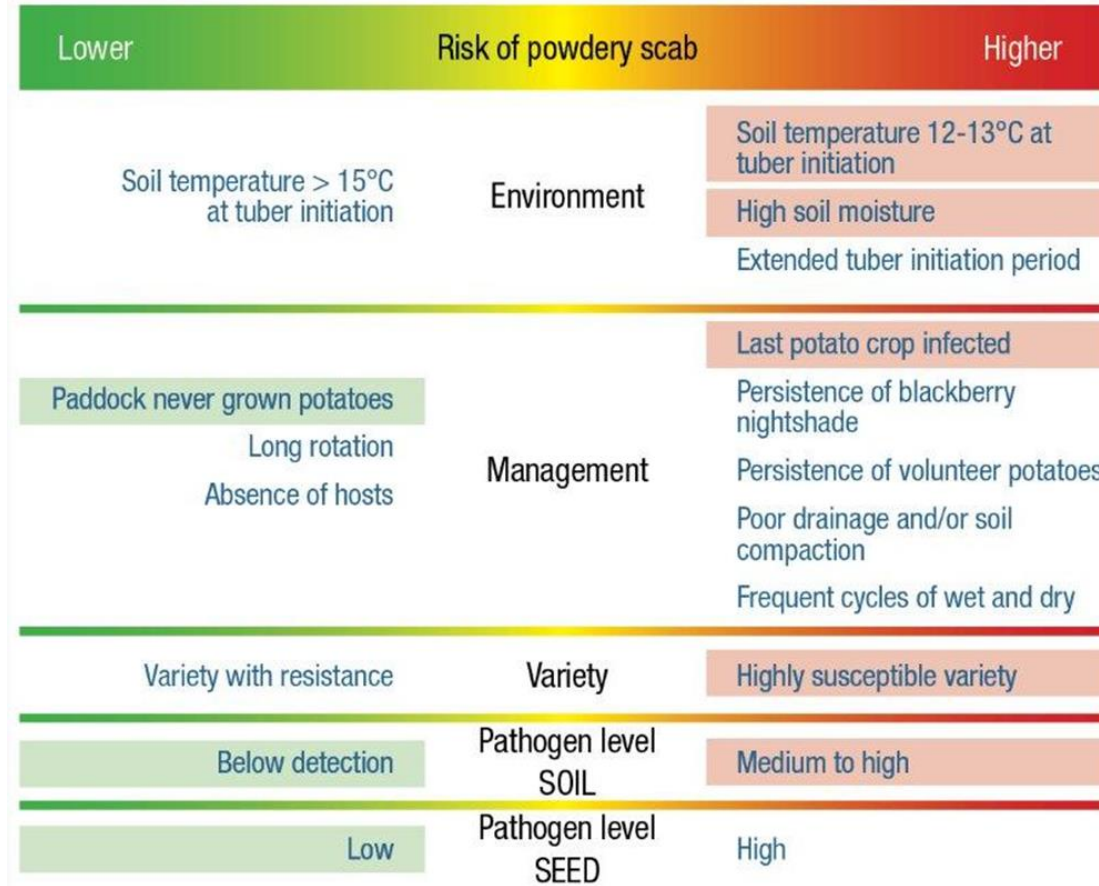
Relating pre-plant soil DNA levels to disease incidence





**Disease: Powdery Scab**  
**Causal Agent:** *Spongospora subterranea*

Powdery Scab



• Greater importance should be placed on highlighted text.

**Understand  
drivers of  
disease**

**Apply  
management  
to reduce risk**

# What do you do with the results?

- Paddock selection
- Crop scheduling
- Varieties
- Paddock and crop rotation
- Cover crop selection
- Soil amendments
- Irrigation
- Crop nutrition
- Fallowing
- Bio-fumigation
- Chemical treatments

**Additional information to inform,  
assess and refine disease  
management strategies**



# To minimise cost soilborne diseases

Growers need to:

- Know which pathogens pose significant risk
- Have management options
- Implement strategy before planting crop

**Management of most soilborne diseases requires that decisions are made and implemented prior to planting**

